

Yale University

Department of Pathology
School of Medicine
108 Lauder Hall
P.O. Box JJJJ
New Haven, Connecticut 06510-8023

Campus address:
108 Lauder Hall
310 Cedar Street
Telephone:
203 785-2719

Re: European Patent EP 0 139 417
Opposition by Chiron Corporation
Genentech Docket: 100/152,233

EXHIBIT G

I, John K. Rose, Ph.D., do declare as follows:

1. On December 14, 1990, I executed a Declaration in support of a reply by Genentech, Inc. (Genentech) to an opposition by Chiron Corporation (Chiron) in connection with Genentech's European Patent No. 0 139 417. A complete copy of my Declaration is attached. I hereby reaffirm all of the statements made in the attached Declaration.

2. I understand that Chiron has now alleged that the opinions stated in my Declaration are not entirely my own. I strongly disagree with these allegations.

3. In the fall of 1990, I was approached by an attorney, Daniel E. Altman, who informed me that he was working on behalf of Genentech in connection with the opposition referred to above. Mr. Altman further informed me that this opposition concerned the work of Genentech researchers, Laurence Lasky and Phillip Berman, regarding their recombinant herpesvirus vaccine. This work related to my primary research interest of the intracellular transport of viral membrane proteins. Accordingly, I was already generally familiar with the work of Drs. Lasky and Berman.

4. Due to my familiarity with these researchers' work, I agreed to review Genentech's patent application, as well as the prior art cited by Chiron in their opposition. After completing this review, I discussed my opinion concerning the nature and predictability of the invention with Mr. Altman. This opinion was based on my review of these documents and on my knowledge of the state of the art at the time this application was filed.

5. Based on our discussion, Mr. Altman prepared a draft Declaration that set forth my qualifications and summarized my opinions. I reviewed this draft and requested that several changes be made. Mr. Altman made the changes and again submitted the draft for my review. I requested that further clarifications be made to the revised draft before finally agreeing that the Declaration accurately set forth my opinions.

6. I understand that Chiron has questioned the statement I made in paragraph 8 of my earlier Declaration concerning the ability of one of ordinary skill in the art to have predicted in August 1983 whether a successful vaccine could have been produced using the process claimed by Genentech. My statement was based on my knowledge of the state of the art at that time. As of August 1983, there had been no previous reports of a vaccine that conferred *in vivo* protection against a pathogen based solely on a truncated, membrane-free

Yale University

Declaration of John K. Rose, Ph.D.
Page -2-

derivative of a viral glycoprotein. (See paragraph 7 of my earlier declaration.) Accordingly, it was not yet known whether such a glycoprotein could give rise to the immune response necessary to confer *in vivo* protection. Moreover, as explained below, it could not have been predicted in August 1983 that such protection would have been conferred.

7. The Gething et al., *Nature*, 300:598-603 (1982) reference cited by Chiron relates to the production of a truncated, membrane-free derivative of haemagglutinin of influenza virus. I was well aware at the time of my earlier declaration that this reference contains an unsubstantiated statement in its penultimate paragraph that the work reported therein could lead to a method for vaccine production. The reference does not contain any other discussion of vaccines. Notwithstanding anything in this Gething et al. reference, one of ordinary skill in the art could not have predicted that a successful vaccine could be produced based on its teachings.

8. As was well known in August 1983, there are a number of significant technical obstacles that must be overcome in order to produce from an isolated glycoprotein a successful vaccine that provides immunoprotection against a pathogen. First, it must be shown that the particular glycoprotein selected actually raises neutralizing antibodies against the pathogen. No such showing is present in the Gething et al. paper. Moreover, it was also well known in August 1983, that *in vivo* protection against a pathogen can often require more than the mere ability to raise antibodies that are neutralizing *in vitro*.

9. Many instances are known in which large numbers of neutralizing antibodies are raised, yet fail to protect the host from pathogenesis. In many instances, pathogens are capable of altering their immunogenic profile so as to escape inactivation by antibodies raised solely against a single glycoprotein. Examples of such pathogens include Equine Infectious Anemia Virus (EIAV), Visna Virus, trypanosomes and HIV-1 (known in 1983 as HTLV-III).

10. Further, as of 1983, it was not known whether a T cell response was required in order to provide protection against many viral pathogens. It was widely thought in 1983 that such cellular immunity was essential to provide an effective vaccine against a viral pathogen, and it was not known if a truncated glycoprotein could evoke such a response.

11. Moreover, those having ordinary skill in the art would not have known whether polyvalent complexes were necessary to provide immunogenicity. For the influenza virus haemagglutinin, Mary Jane Gething, the lead author of the Gething et al. reference, was quoted in Zoller et al., *Bio/Technology*, pp.146-147 (April 1983), as stating that "[c]learly, one needs polyvalent complexes [of HA molecules] for immunogenicity." However, unexpectedly, Lasky and Berman demonstrated that a single truncated, membrane-free derivative of herpes simplex virus (HSV) gC or gD glycoprotein could successfully protect animals from infection by HSV.

Yale University

Declaration of John K. Rose, Ph.D.

Page -3-

12. Thus, as of the effective filing date of this application, it could by no means be predicted that a successful vaccine could be produced based solely on a single truncated, membrane free derivative of a viral glycoprotein. One of ordinary skill in the art at the relevant date could not have predicted whether such a glycoprotein would elicit neutralizing antibodies that would be effective in preventing pathogenesis, whether the antibodies alone without a cellular immune component could be effective, nor whether a complex of glycoproteins was required for immunogenicity. Thus, in August 1983, one of ordinary skill in the art would not have had sufficient information to predict Lasky's and Berman's successful results.

13. In paragraph 9 of my earlier declaration, I stated that once Drs. Lasky and Berman demonstrated the successful production of a vaccine in their HSV model that a reasonable expectation arose that the system would be successful with other viral pathogens. This expectation arose because the successful results produced in the HSV model demonstrated that all of the technical challenges to successful vaccine production had been overcome. In other words, Lasky's and Berman's successful production of a vaccine effective against a viral pathogen based on a truncated, membrane-free viral glycoprotein showed that such a glycoprotein could elicit neutralizing antibodies and that such antibodies could be raised against a single glycoprotein. Lasky's and Berman's successful work further showed that such antibodies could be effective in preventing pathogenesis, and that any necessary cellular immune component must have been generated. The work of Gething et al. showed nothing with respect to these technical obstacles. Thus, the Gething et al. work did not give rise to any expectation of success.

14. I understand that I was quoted in the Zoller et al. article referred to above, as stating that the idea that removal of the anchor sequence could convert these proteins from a membrane-bound to a secretory form was "pretty much obvious to everybody." This is certainly what was expected at the time the Gething et al. reference referred to above was published. However, as explained above, the expectation that the proteins could be secreted suggests nothing regarding whether the secreted proteins would provide an effective vaccine.

15. I declare that all statements made herein of my own knowledge are true, and that all statements made upon information and belief are believed to be true, and, further, that these statements were made with the knowledge that willful, false statements and the like so made are punishable by fine or imprisonment, or both, and that willful, false statements may jeopardize the validity of the patent.

Dated: Dec. 20, 1993

By: John K. Rose
John K. Rose, Ph.D.

Publications:

1. Rose, J.K., Mosteller, R.D. and Yanofsky, C. 1970. Tryptophan messenger RNA elongation rates and steady state levels of tryptophan operon enzymes under various growth conditions. *J. Mol. Biol.* 51:541-550.
2. Mosteller, R.D., Rose, J.K. and Yanofsky, C. 1970. Transcription initiation and degradation of trp mRNA. *Cold Spring Harbor Symp. Quant. Biol.* 35:461-466.
3. Rose, J.K. and Yanofsky, C. 1971. Transcription of the operator proximal and distal ends of the tryptophan operon: evidence that trp E and trp A and the delimiting structural genes. *J. Bacteriol.* 108:615-618.
4. Rose, J.K. and Yanofsky, C. 1972. Metabolic regulation of the tryptophan operon of *Escherichia coli*: Repressor-independent regulation of transcription initiation frequency. *J. Mol. Biol.* 69:103-118.
5. Rose, J.K., Squires, C.L., Yanofsky, C., Yang, H.-L. and Zubay, G. 1973. Regulation of *in vitro* transcription of the tryptophan operon by purified RNA polymerase in the presence of partially purified repressor and tryptophan. *Nature New Biol.* 245:133-137.
6. Squires, C.L., Rose, J.K., Yanofsky, C., Yang, H.-L. and Zubay, G. 1973. Tryptophanyl-tRNA and tryptophanyl-tRNA synthetase are not required for *in vitro* repression of the tryptophan operon. *Nature New Biol.* 245:131-133.
7. Rose, J.K. and Yanofsky, C. 1974. Interaction of the operator of the tryptophan operon with repressor. *Proc. Natl. Acad. Sci. USA* 71: 3134-3138.
8. Rose, J.K. and Knipe, D. 1975. Nucleotide sequence complexities, molecular weights and poly(A) content of the vesicular stomatitis virus mRNA species. *J. Virol.* 15:994-1003.
9. Knipe, D., Rose, J.K. and Lodish, H.F. 1975. Translation of individual species of vesicular stomatitis viral mRNA. *J. Virol.* 15:1004-1011.
10. Rose, J.K. 1975. Heterogeneous 5'-terminal structures occur on vesicular stomatitis virus mRNAs. *J. Biol. Chem.* 250:8098-8104.
11. Hewlett, M.J., Rose, J.K. and Baltimore, D. 1976. 5'-terminal structure of poliovirus polyribosomal RNA is pUp. *Proc. Natl. Acad. Sci. USA* 73:327-330.
12. Rose, J.K. and Lodish, H.F. 1976. Translation *in vitro* of vesicular stomatitis virus mRNA lacking 5'-terminal 7-methylguanosine. *Nature* 262:32-37.
13. Rose, J.K., Hasletine, W.A. and Baltimore, D. 1976. The 5' terminus of Moloney murine leukemia virus 35S RNA is m7G5'ppp5'GmpCp. *J. Virol.* 20:324-329.
14. Lodish, H.F. and Rose, J.K. 1977. Relative importance of 7-methylguanosine in ribosome binding and translation of vesicular stomatitis virus mRNA in wheat germ and reticulocyte cell-free systems. *J. Biol. Chem.* 252:1181-1188.
15. Rose, J.K., Lodish, H.F. and Brock, M.L. 1977. Giant heterogeneous poly(A) on vesicular stomatitis virus mRNA synthesized *in vitro* in the presence of S-adenosylhomocysteine. *J. Virol.* 21:683-693.
16. Freeman, G.J., Rose, J.K., Clinton, G.M. and Huang, A.S. 1977. RNA synthesis of vesicular stomatitis virus VII. Complete separation of the messenger RNAs of vesicular stomatitis virus by duplex formation. *J. Virol.* 21:1094-1104.
17. Rose, J.K. 1977. Nucleotide sequences of ribosome recognition sites in messenger RNAs of vesicular stomatitis virus. *Proc. Natl. Acad. Sci. USA* 74:3672-3676.
18. Pettersson, R.F., Flanagan, J.B., Rose, J.K. and Baltimore, D. 1977. 5'-terminal nucleotide sequences of polio virus polyribosomal RNA and virion RNA are identical. *Nature (London)* 268:270-272.

19. Rose, J.K. 1977. Ribosome recognition sites in vesicular stomatitis virus mRNA. In, Negative Strand Viruses and the Host Cell (B.W.J. Mahy and R.D. Barry, eds.), Academic Press, New York, pp. 47-61.
20. Baltimore, D., Pettersson, R.F., Flanegan, J.B., Hewlett, M.J., Rose, J.K. and Ambros, V. 1978. New structures in viral RNA: Non-covalent circles and covalently-linked protein. In, Perspectives in Virology (M. Pollard, ed.), pp. 110-115.
21. Rose, J.K. 1978. Complete sequences of the ribosome recognition sites in vesicular stomatitis virus mRNAs: Recognition by the 40S and 80S complexes. *Cell* 14:345-353.
22. Rose, J.K., Trachsel, H., Leong, K. and Baltimore, D. 1978. Inhibition of translation by poliovirus: Inactivation of a specific initiation factor. *Proc. Natl. Acad. Sci. USA* 75:2732-2736.
23. Rose, J.K. and Iverson, L. 1979. Nucleotide sequences from the 3'-ends of vesicular stomatitis virus mRNAs as determined from cloned DNA. *J. Virol.* 32:404-411.
24. Rose, J.K. 1980. Ribosome recognition and translation of vesicular stomatitis virus messenger RNA. In, Rhabdoviruses (D.H.L. Bishop, ed.), CRC Press, Boca Raton, Florida, pp. 51-60.
25. Trachsel, H., Sonnenberg, N., Shatkin, A.J., Rose, J.K., Leong, K., Bergmann, J.E., Gordon, J. and Baltimore, D. 1980. Purification of a factor that restores translation of vesicular stomatitis virus mRNA in extracts from poliovirus-infected HeLa cells. *Proc. Natl. Acad. Sci. USA* 77:770-774.
26. Rose, J.K. 1980. Complete intergenic and flanking gene sequences from the genome of vesicular stomatitis virus. *Cell* 19:415-421.
27. Rose, J.K., Welch, W.J., Sefton, B.M., Esch, F.S. and Ling, N.C. 1980. Vesicular stomatitis virus glycoprotein is anchored in the viral membrane by a hydrophobic domain near the COOH-terminus. *Proc. Natl. Acad. Sci. USA* 77:3884-3888.
28. Rose, J.K., Welch, W.J., Sefton, B.M. and Iverson, L.E. 1980. Analysis of VSV glycoprotein structure and genome structure using cloned DNA. In, Animal Virus Genetics (B.N. Fields, R. Jaenisch and C.F. Fox, eds.), Academic Press, New York.
29. Iverson, L.E. and Rose, J.K. 1981. Localized attenuation and discontinuous synthesis during vesicular stomatitis virus transcription. *Cell* 23:477-484.
30. Rose, J.K., Iverson, L.E., Gallione, C.J. and Greene, J.R. 1981. Vesicular stomatitis virus gene structure and transcription attenuation. In, Negative Strand Viruses (D.H.L. Bishop and R. Compans, eds.), Elsevier, North Holland, pp. 713-721.
31. Rose, J.K. and Gallione, C.J. 1981. Nucleotide sequences of the mRNA's encoding the vesicular stomatitis virus G and M proteins determined from cDNA clones containing the complete coding regions. *J. Virol.* 39:519-528.
32. Gallione, C., Greene, J., Iverson, L. and Rose, J.K. 1981. Nucleotide sequences of the mRNA's encoding the vesicular stomatitis virus N and NS proteins. *J. Virol.* 39:529-535.
33. Rose, J.K. and Shafferman, A. 1981. Conditional expression of the vesicular stomatitis virus glycoprotein gene in *Escherichia coli*. *Proc. Natl. Acad. Sci. USA* 78:6670-6674.
34. Rose, J.K., Doolittle, R., Anilionis, A., Curtis, P., Wunner, W. 1982. Homology between the glycoproteins of vesicular stomatitis virus and rabies virus. *J. Virol.* 43:361-364.
35. Rose, J.K. and Bergmann, J.E. 1982. Expression from cloned cDNA of cell-surface and secreted forms of the glycoprotein of vesicular stomatitis virus in eucaryotic cells. *Cell* 30:753-762.
36. Iverson, L.E. and Rose, J.K. 1982. Sequential synthesis of 5'-proximal vesicular stomatitis virus mRNA sequences. *J. Virol.* 44: 356-365.

37. Gallione, C.J. and Rose, J.K. 1983. Nucleotide sequence of a cDNA clone encoding the entire glycoprotein from the New Jersey serotype of vesicular stomatitis virus. *J. Virol.* 46:162-169.
38. Rose, J.K. and Bergmann, J.E. 1983. Altered cytoplasmic domains affect intracellular transport of the vesicular stomatitis virus glycoprotein. *Cell* 34:513-524.
39. Florkiewicz, R.Z., Smith, A., Bergmann, J.E. and Rose, J.K. 1983. Isolation of stable mouse cell lines that express cell surface and secreted forms of the vesicular stomatitis virus glycoprotein. *J. Cell Biol.* 97:1381-1388.
40. Rose, J.K., Adams, G.A. and Gallione, C.J. 1984. The presence of cysteine in the cytoplasmic domain of the vesicular stomatitis virus glycoprotein is required for palmitate addition. *Proc. Natl. Acad. Sci. USA* 81:2050-2054.
41. Florkiewicz, R.Z. and Rose, J.K. 1984. A cell line expressing the vesicular stomatitis virus glycoprotein fuses at low pH. *Science* 225:721-723.
42. Guan, J.-L. and Rose, J.K. 1984. Conversion of a secretory protein into a transmembrane protein results in its transport to the Golgi complex but not to the cell surface. *Cell* 37:779-787.
43. Gallione, C.J. and Rose, J.K. 1985. A single amino acid substitution in a hydrophobic domain causes temperature-sensitive cell-surface transport of a mutant viral glycoprotein. *J. Virol.* 54:374-382.
44. Moller, J.R., Rager-Zisman, B., Phuc-Cahn, Q., Schattner, A., Panush, D., Rose, J.K. and Bloom, B.R. 1985. NK cell recognition of target cells expressing different antigens of vesicular stomatitis virus. *Proc. Natl. Acad. Sci. U.S.A.* 82:2456-2459.
45. Yilma, T., Mackett, M., Rose, J. and Moss, B., 1985. Vaccinia virus recombinants expressing vesicular stomatitis genes immunize mice and cattle. *Science* 277:433-435.
46. Adams, G.A. and Rose, J.K. 1985. Incorporation of a charged amino acid into the membrane spanning domain blocks cell-surface transport but not membrane-anchoring of a viral glycoprotein. *Mol. Cell Biol.* 5:1442-1448.
47. Rose, J., Adams, G., Guan, J.-L., Machamer, C. and Puddington, L. 1985. Redesigning transport signals in membrane and secretory proteins. (*In*, Protein Transport and Secretion, pp. 7-85; ed. Mary-Jane Gething). Cold Spring Harbor Laboratory Press, 1985.
48. Adams, G.A. and Rose, J.K. 1985. Structural requirements of a membrane spanning domain for protein anchoring and cell surface transport. *Cell* 41:1007-1015.
49. Machamer, C.E., Florkiewicz, R.Z. and Rose, J.K. 1985. A single N-linked oligosaccharide at either of the two normal sites is sufficient for transport of the vesicular stomatitis virus G protein to the cell surface. *Mol. Cell Biol.* 5:3074-3083.
50. Guan, J.-L., Machamer, C.E. and Rose, J.K. 1985. Glycosylation allows cell-surface transport of an anchored secretory protein. *Cell* 42:489-496.
51. Rose, J.K. and Schubert, M. 1987. Rhabdovirus Genomes and Their Products. *In*, The Viruses, (eds. R. Wagner and H. Frankel-Conrat) Plenum Press, pp. 129-159.
52. Machamer, C.E., Guan, J.-L., Florkiewicz, R.Z. and Rose, J.K. 1986. Role of N-linked glycosylation in intracellular transport of transmembrane proteins. *Microbiology* (Wash. D.C.) 1986:292-296.
53. Woodgett, C. and Rose, J.K. 1986. Amino-terminal mutation of the vesicular stomatitis virus glycoprotein does not affect its fusion activity. *J. Virol.* 59:486-489.

54. Puddington, L., Machamer, C.E. and Rose, J.K. 1986. Cytoplasmic domains of cellular and viral integral membrane proteins substitute for the cytoplasmic domain of the vesicular stomatitis virus glycoprotein in transport to the plasma membrane. *J. Cell Biol.* 102:2147-2157.
55. Puddington, L., Bevan, M.J., Rose, J.K. and Lefrançois, L. 1986. N protein is the predominant antigen recognized by vesicular stomatitis virus-specific cytotoxic T Cells. *J. Virol.* 60: 708-717.
56. Puddington, L., Woodgett, C. and Rose, J.K. 1987. Replacement of the cytoplasmic domain alters sorting of a viral glycoprotein in polarized cells. *Proc. Natl. Acad. Sci., USA* 84:2756-2760.
57. Scullion, B.F., Hou, Y., Puddington, L., Rose, J.K. and Jacobson, K. 1987. Effects of mutations in three domains of the vesicular stomatitis viral glycoprotein on its lateral diffusion in the plasma membrane. *J. Cell Biol.* 105:69-75.
58. Rottier, P.J.M. and Rose, J.K. 1987. Coronavirus E1 glycoprotein expressed from cloned cDNA localizes in the Golgi region. *J. Virol.* 61:2042-2045.
59. Rottier, P.J.M., Florkiewicz, R.Z., Shaw, A.S. and Rose, J.K. 1987. An internalized amino-terminal signal sequence retains full activity in vivo but not in vitro. *J. Biol. Chem.* 262:8889-8895.
60. Machamer, C.E. and Rose, J.K. 1987. A specific transmembrane domain of a coronavirus E1 glycoprotein is required for its retention in the Golgi region. *J. Cell Biol.* 105:1205-1214.
61. Kupfer, A., Kronebusch, P.J., Rose, J.K. and Singer, S.J. 1987. A critical role for the polarization of membrane recycling in cell motility. *Cell Motility and the Cytoskeleton.* 8:182-189.
62. Guan, J.-L., Cao, H. and Rose, J.K. 1988. Cell-surface expression of a membrane-anchored form of the human chorionic gonadotropin alpha subunit. *J. Biol. Chem.* 263:5306-5313.
63. Fukuda, M., Guan, J.-L. and Rose, J.K. 1988. A membrane-anchored form but not the secretory form of human chorionic gonadotropin alpha chain acquires polygalactosaminoglycan. *J. Biol. Chem.* 263:5314-5318.
64. Machamer, C.E. and Rose, J.K. 1988. Influence of new glycosylation sites on expression of the vesicular stomatitis virus G protein at the plasma membrane. *J. Biol. Chem.* 263:5948-5954.
65. Machamer, C.E. and Rose, J.K. 1988. Vesicular stomatitis virus G proteins with altered glycosylation sites display temperature-sensitive intracellular transport and are subject to aberrant intermolecular disulfide bonding. *J. Biol. Chem.* 263:5955-5960.
66. Doms, R.W., Ruusala, A., Machamer, C., Helenius, J., Helenius, A. and Rose, J.K. 1988. Differential effects of mutations in three domains on folding, quaternary structure, and intracellular transport of vesicular stomatitis virus G protein. *J. Cell Biol.* 107:89-99.
67. Guan, J.-L., Ruusala, A., Cao, H. and Rose, J.K. 1988. Effects of altered cytoplasmic domains on transport of the vesicular stomatitis virus glycoprotein are transferable to other proteins. *Mol. Cell Biol.* 8:2869-2874.
68. Shaw, A.S., Rottier, P.J.M. and Rose, J.K. 1988. Evidence for the loop model of signal sequence insertion into the endoplasmic reticulum. *Proc. Natl. Acad. Sci. USA* 85:7592-7596.
69. Rose, J.K. and Doms, R.W. 1988. Regulation of protein export from the endoplasmic reticulum. *Ann. Rev. Cell Biol.* 4:257-288.

70. Whitt, M., Chong, L. and Rose, J.K. 1989. Glycoprotein cytoplasmic domain sequences required for rescue of the VSV glycoprotein mutant. *J. Virol.* 63:3569-3578.
71. Pitta, A.M., Rose, J.K., and Machamer, C.E. A single amino acid substitution eliminates the stringent carbohydrate requirement for the intracellular transport of a viral glycoprotein. *J. Virol.* 63:3801-3809.
72. Shaw, A., Amrein, K., Hammond, C., Stern, D.F., Sefton, B.M. and Rose, J.K. 1989. The *Ick* tyrosine protein kinase interacts with the cytoplasmic tail of the CD4 glycoprotein through its unique amino-terminal domain. *Cell* 59:627-636.
73. Brown, D.A., Crise, B., and Rose, J.K. 1989. Mechanism of membrane anchoring affects polarized expression of two proteins in MDCK cells. *Science* 245:1499-1501.
74. Zagouras, P., and Rose, J.K. 1989. Carboxy-terminal SEKDEL sequences retard but do not retain two secretory proteins in the endoplasmic reticulum. *J. Cell Biol.* 109:2633-2640.
75. Crise, B., Ruusala, A., Zagouras, P., Shaw, A., and Rose, J.K. 1989. Oligomerization of glycolipid-anchored and soluble forms of the vesicular stomatitis virus glycoprotein. *J. Virol.* 63:5328-5333.
76. Miettinen, H.M., Rose, J.K., and Mellman, I. 1989. Fc receptor isoforms exhibit distinct abilities for coated pit localization as a result of cytoplasmic domain heterogeneity. *Cell* 58:317-327.
77. Shaw AS, J Chalupny, JA Whitney, C Hammond, KE Amrein, P Kavathas, BA Sefton and JK Rose. 1990. Short related sequences in the cytoplasmic domains of CD4 and CD8 mediate binding to the amino-terminal domain of the p56^{lck} tyrosine protein kinase. *Mol. Cell Biol.* 10:1853-1862.
78. Machamer CE, RW Doms, DG Bole, A Helenius and JK Rose. 1990. BiP recognizes incompletely disulfide-bonded forms of vesicular stomatitis virus G protein. *J. Biol. Chem.* 265:6879-6883.
79. Machamer CE, SA Mentone, JK Rose and MG Farquhar. 1990. The E1 glycoprotein of an avian coronavirus is targeted to the *cis* Golgi complex. *Proc. Natl. Acad. Sci. USA* 87:6944-6948.
80. Buonocore L, and Rose JK. 1990. Prevention of HIV-1 glycoprotein transport by soluble CD4 retained in the endoplasmic reticulum. *Nature* 345:625-628.
81. Whitt MA, Zagouras P, Crise B, & Rose JK. 1990. A fusion-defective mutant of the vesicular stomatitis virus glycoprotein. *J. Virol.* 64:4907-4913.
82. Rose JK, Buonocore L, & Whitt MA. 1990. Cationic liposome reagents for efficient uptake of DNA and transient expression in the cytoplasm of animal cells. *Biotechniques* {submitted}
83. Crise, B., Buonocore, L., and Rose, J.K. 1990. CD4 is retained in the endoplasmic reticulum by the HIV-1 glycoprotein precursor. *J. Virol.* 64:5585-5593.

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- BLACK BORDERS**
- IMAGE CUT OFF AT TOP, BOTTOM OR SIDES**
- FADED TEXT OR DRAWING**
- BLURRED OR ILLEGIBLE TEXT OR DRAWING**
- SKEWED/SLANTED IMAGES**
- COLOR OR BLACK AND WHITE PHOTOGRAPHS**
- GRAY SCALE DOCUMENTS**
- LINES OR MARKS ON ORIGINAL DOCUMENT**
- REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY**
- OTHER:** _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.